


Treatment of critical-sized bone defects: clinical and tissue engineering perspectives

Erika Roddy¹  · Malcolm R. DeBaun² · Adam Daoud-Gray³ · Yunzhi P. Yang^{4,5,6} · Michael J. Gardner²

Received: 2 May 2017 / Accepted: 8 October 2017 / Published online: 28 October 2017
© Springer-Verlag France SAS 2017

Abstract Critical-sized bone defects are defined as those that will not heal spontaneously within a patient's lifetime. Current treatment options include vascularized bone grafts, distraction osteogenesis, and the induced membrane technique. The induced membrane technique is an increasingly utilized method with favorable results including high rates of union. Tissue engineering holds promise in the treatment of large bone defects due to advancement of stem cell biology, novel biomaterials, and 3D bioprinting. In this review, we provide an overview of the current operative treatment strategies of critical-sized bone defects as well as the current state of tissue engineering for such defects.

Keywords Critical bone defects · Bone tissue engineering · Bone healing · Fracture healing

Introduction

The management of critical-sized bone defects remains a major clinical orthopedic challenge. Critical-sized bone defects are technically defined as those that will not heal spontaneously during the patient's lifetime [121]. Bone loss greater than 2 times the diameter of the long bone diaphysis is unlikely to result in union despite appropriate stabilization methods [56, 85]. Large bone defects can be secondary to bone loss from trauma, infection, tumor resection, or developmental deformities. The overall incidence of critical-sized bone defects is low. For instance, in one 10-year fracture registry, only 0.4% of all fractures at a level-1 trauma center were complicated by significant bone loss [63].

The condition of the soft tissues surrounding the bone defect is a major contributory factor to bone healing. Unlike other tissues, bone can regenerate and repair itself through the process of primary or secondary healing. However, this process is dependent on adequate vascularity, and avascularity is a major contributor to the pathogenesis of critical-sized defects [78, 129]. The majority of fractures heal by secondary healing. This process depends on osteogenesis, osteoinduction, and osteoconduction [30]. Specifically, mesenchymal stem cells (present in the bone marrow, granulation tissue, periosteum, surrounding soft tissues, and blood vessels) provide the osteoprogenitor cells that differentiate into osteoblasts and osteoclasts, while osteoinductive factors stimulate this differentiation. Osteoinductive factors are delivered by vasculature to the site of the fracture and include pro-inflammatory cytokines, growth factors such as those in the TGF- β superfamily (e.g., bone morphogenic proteins) and angiogenic factors such as VEGF. Finally, an osteoconductive scaffold, which permits bone growth onto its surface, is also necessary. In native fracture healing, this

✉ Michael J. Gardner
michaelgardner@stanford.edu

¹ School of Medicine, University of California, San Francisco (UCSF), 513 Parnassus Ave, San Francisco, CA 94143, USA

² Department of Orthopaedic Surgery, Stanford University, 300 Pasteur Drive, Stanford, CA 94305, USA

³ School of Medicine, Stanford University School of Medicine, 300 Pasteur Drive, Stanford, CA 94305, USA

⁴ Department of Orthopedic Surgery, Stanford University, 300 Pasteur Drive, Stanford, CA 94305, USA

⁵ Department of Bioengineering, Stanford University, 318 Campus Drive, Stanford, CA 94305, USA

⁶ Department of Materials Science and Engineering, Stanford University, 300 Pasteur Drive, Stanford, CA 94305, USA

scaffold is first provided by the hematoma and then by the cartilage callus [30].

Host factors can also adversely affect bone healing. Comorbid metabolic and systemic disorders such as diabetes, obesity, inflammatory arthritis, malnutrition, peripheral vascular disease, and hypothyroidism, as well as certain drugs such as NSAIDs, steroids, phenytoin, ciprofloxacin, and anticoagulants are known to increase healing time and risk of nonunion [2, 23, 39, 48, 51, 55]. Specifically, diabetes is associated with up to three times the risk of nonunion and a doubling of the time to healing [65, 75]. Pro-inflammatory cytokines associated with systemic inflammatory diseases including diabetes, obesity, and inflammatory arthritis are thought to activate osteoclasts and therefore create an imbalance between bone resorption and formation [23]. Inflammatory diseases are also associated with reduced expression of osteoinductive factors [114]. Finally, patient behaviors including smoking and alcohol abuse are also associated with significantly delayed union and increased risk of nonunion [39]. Alcohol inhibits cell proliferation [21]. Smoking is associated with atherosclerosis and reduced blood supply [104, 107]. Further, nicotine inhibits vascular ingrowth and early revascularization of bone and diminishes osteoblast function [26, 116]. Advanced age, venous stasis, burns, radiation, and obesity are additional predisposing factors [34, 48, 66].

There is a paucity of evidence to guide clinical treatment strategies for critical-sized bone defects, with no controlled studies comparing different techniques. Historically, large segmental defects have had a poor prognosis, often requiring amputation. Current therapeutic approaches include bone grafting, distraction osteogenesis, and the induced membrane (“Masquelet”) technique. Although case series and retrospective reviews have reported relatively high rates of union following these techniques, each is associated with its own complications. Therefore, much recent research has been devoted to tissue engineering strategies that can provide all three of the factors deemed essential for bone healing: osteoconductive scaffold, growth factors for osteoinduction, and cells with osteogenic potential. The purpose of this review is to provide an overview of the current operative treatment strategies of critical-sized bone defects as well as the current state of tissue engineering for such defects.

Nonstructural bone grafts: cancellous autograft

Nonstructural bone grafts include autologous cancellous grafts, allografts in the form of mineralized allografts or demineralized bone matrices (DBMs), and bone graft substitutes including calcium sulfate, calcium phosphate, hydroxyapatite, and tricalcium phosphate. Cancellous autograft remains the gold standard void filler for

the treatment of bone loss during nonunion treatment. It provides osteoconductive (scaffold, allowing ingrowth of osteoblasts), osteoinductive (bone morphogenetic proteins and other growth factors which signal the differentiation of mesenchymal cells along the osteogenic lineage) and osteogenic (mesenchymal stem cells) factors [13]. Additionally, it is nonimmunogenic and does not carry the risk of transmissible infections [13]. However, for large segmental defects, the morbidity associated with harvesting the amount of graft required to fill the defect is substantial; additionally, the amount of graft that can be obtained is finite. Historically, autograft has been harvested from the anterior and/or posterior iliac crest. For iliac crest bone grafting, the rate of complications has been reported to be in the range of 20% [29, 43], with pain at the harvest site reported in 18% of patients at 2 years [29]. Recently, the Reamer/Irrigator/Aspirator (RIA) device has offered a new method of obtaining bone graft. The RIA addresses the reaming-related problems of thermal necrosis and embolism of marrow contents through simultaneous irrigation of the canal and suction of reaming debris [53, 103]. This same debris has recently been utilized as bone graft. In fact, RIA reamings contain multipotent stem cells as well as high levels of growth factors [120]. Additionally, the volume of bone graft obtained from RIA is significantly larger (25–90 cm³) than that obtained from anterior iliac crest harvesting (5–72 cm³), although comparable to that obtained from posterior crest harvesting (25–88 cm³) [29]. Importantly, a review of 233 patients undergoing RIA showed only a 6% complication rate [29]. Complications reported after RIA include iatrogenic fracture, violation of the knee joint, and heterotopic ossification [24, 76, 110].

Over the past two decades, surgeons have begun to utilize bone morphogenetic proteins (BMPs) as an alternative to, or in combination with, autologous bone grafts. BMPs are growth factors important in the formation of bone and cartilage as well as bone repair. BMP-2 and BMP-7 are the most clinically relevant of this group. In 2001, recombinant BMP-7 (OP-1) became FDA-approved for the treatment of tibial nonunions after a randomized control trial showed no difference in the rate of union in patients treated with BMP-7 plus intramedullary (IM) rod versus fresh autograft plus IM rod for tibial nonunion [36]. In 2002, BMP-2 was FDA-approved for the treatment of acute open tibial fractures after a randomized control trial comparing BMP-7 plus IM nail versus IM nail alone found a decreased rate of secondary intervention in the experimental group [46]. Since then, off-label use of these molecules has been adopted by many surgeons for treatment of nonunion sites at other locations [15, 35, 59]. Unfortunately, a Cochrane review recently found that the rate of union is not different for nonunions treated with BMPs versus autologous graft [RR 1.02 (95% CI 0.9 to 1.15)] [38].

Moreover, although BMPs have strong osteoinductive properties, surgeons need to be cognizant that they lack osteoconductive and osteogenic factors. This is particularly relevant for the use of BMPs in the treatment of critical-sized bone defects—if used for such applications, they must be combined with autologous bone graft or another scaffold (the current clinical delivery system is a collagen sponge) and supply of osteoprogenitor cells. More studies need to be performed evaluating the role of BMP plus autologous graft versus autologous graft alone.

Bone marrow aspirate represents another option for obtaining osteoprogenitor cells, as well as osteoinductive factors. However, it, like BMPs, has no osteoconductive properties. Some studies have reported union rates from 75 to 88% using bone marrow aspirate for the treatment of nonunions; however, they had no control groups [42, 52]. Moreover, it cannot be used alone for the treatment of critical-size defects.

Although autologous bone graft represents the gold standard for nonunions and has osteoinductive, osteoconductive, and osteogenic properties, its lack of structural support makes such grafts, with or without the addition of BMPs, less desirable for larger defects. The graft becomes partially resorbed and revascularization, and bridging occurs by creeping substitution with cells migrating from the well-perfused section into the graft, both of which lead to weakness of the reconstructed segment and increase the risk of iterative fractures [82]. Currently, cancellous autograft is recommended for voids less than 5 cm, with well-vascularized, healthy recipient sites, and that do not require structural integrity from the graft [43]. Therefore, for segmental bone defects larger than 5 cm, other therapeutic options are necessary. These options include allografts, vascularized fibular grafts, distraction osteogenesis and bone transport, and the induced membrane technique.

Nonstructural bone grafts: cancellous allograft

The benefits of cancellous allograft include availability of graft in the desired quantity, avoidance of donor site morbidity, the ability of the graft to provide structural support, and a relatively straightforward surgical technique [91, 92]. However, cancellous allografts are also not vascularized and therefore have limited ability to integrate with host bone. Additionally, they have no osteogenic potential as mesenchymal stem cells, osteoblasts, or osteoclasts are unable to survive under conditions of low oxygen tension [40]. Allografts are also associated with an increased risk of disease transmission as well as immunogenic response [93, 130]. Studies utilizing cadaveric allografts in critical-sized defects have demonstrated frequent complications including nonunion, fracture, and infection [5, 33, 37]. In one study of

oncologic patients, the rate of nonunion was 75%, the rate of fracture was 13%, and the total rate of infection was 16% at an average of 5 years of follow-up [17]. However, the overall rate of allograft survival was 81%, consistent with most reports in the literature [5, 33, 37]. In general, cadaveric allograft is utilized mostly for massive oncologic-related defects in association with endoprosthetic repair and is not commonly utilized for defects arising in the setting of trauma or infection. Recently, orthopedic oncologists have begun to combine massive bone allograft and autologous vascularized fibular grafts, with promising results including improved rates of union (over 70%) [54, 68, 111].

Structural bone grafts: free vascularized fibular grafts

Vascularized autogenous cortical bone grafts, by virtue of maintaining their blood supply, allow the cells contained in these grafts including mesenchymal stem cells, osteoblasts, and osteoclasts to remain viable [128]. This preserves bone-remodeling ability even when the blood supply is inadequate at the recipient site. By contrast, conventional non-vascularized bone grafts are incapable of remodeling unless the recipient site has adequate blood supply. Together with the ability to provide structural support, vascularized grafts are ideal for the treatment of segmental long bone defects.

The size of the fibula matches that of both the radius and ulna and thus can anatomically match forearm deficits. For larger long bones such as the femur or proximal tibia, the fibular graft can be inserted into the medullary canal singularly, with another free fibular graft, or combined with allograft [14]. It may also be cut transversely and folded to match the width of the recipient bone site while preserving the vascular pedicle. This is particularly useful when considering the cross-sectional area of the femur compared to the fibula and can reduce the risk of stress fracture after weight bearing. Another advantage of the fibula is that it tends to hypertrophy in response to microscopic stress fractures, and studies have reported rapid hypertrophy when free fibular grafts are used for tibial or femoral defects [18, 27]. Rates of union after vascularized free fibular grafting range from 70 to 100% in retrospective studies of critical-sized defects secondary to a variety of causes including trauma, osteomyelitis, and tumor resection [28, 32, 67, 89, 96, 137, 139, 141]. Mean time to union is approximately 6 months [28, 89, 96, 139]. The rate of return to weight bearing and adequate functionality, when reported, is also generally high: over 96% in one study [71].

However, the rate of fractures after free fibular grafting in the lower extremities may range from 15 to over 40% [27, 28, 67, 89]. These fractures are thought to occur due to excessive mechanical stress or misalignment. For this

reasons, some authors recommend fibular grafts be used predominantly for the upper extremities alone [113, 128, 137]. Others have found lower rates of fracture by using conventional plates, external fixators, or locked plates in conjunction with the fibular graft, as well as implementing delayed weight bearing [70]. Still others hypothesize that stress fractures themselves are desirable because they induce bony hypertrophy through activation of bone remodeling pathways [108, 139].

Other reported complications include donor site morbidity, failed anastomosis, microvascular thrombosis, infection, and progressive deformities [6, 90, 133]. A systemic review of complications of free fibula grafts found an overall incidence of early donor site complications (including infection, dehiscence, delayed wound healing) of 9.9% for wounds closed primarily, and 19.0% for wounds requiring skin graft closure [72]. Late morbidities included chronic pain (6.5%), gait abnormality (3.9%), ankle instability (5.8%), limited range of motion (11.5%), and sensory deficit (7.0%) [72]. Overall, disadvantages of the fibular autograft include donor site morbidity, increased operative time, risk of fracture particularly in the lower extremities, and a challenging microsurgical technique. However, the fibula graft may still be useful in cases of segmental bone loss in the upper extremity, especially those with associated soft tissue loss [49, 94, 113, 128, 137].

Distraction osteogenesis and bone transport

Ilizarov first reported on the technique of distraction osteogenesis in the 1950s [9]. This technique utilizes the bone's natural capacity for regeneration under tension. A corticotomy is made in healthy bone, usually metaphyseal, at a distance from the defect site to create a free segment of living bone [9]. This is followed by distraction of the free segment toward the defect site, the future docking site, with bone production occurring *de novo* between the two corticotomy surfaces. The process of bone formation in the distraction gap is histologically similar to intramembranous ossification [10, 11]. Distraction-compression transport is achieved mechanically by attaching the segmental fragments to a circular external fixator with tensioned wires, which allows for distraction at the corticotomy site and eventually compression at the docking site.

The Ilizarov technique is divided into three periods. The first is the latency period, which is the time from corticotomy to the beginning of distraction during which callus is formed, usually 5–7 days. Next is the distraction period. A distraction rate of 1 mm per day has been shown to minimize the rate of premature consolidation, yet does not outpace the speed of vascular ingrowth [12]. Finally, once the bone fragment closes the defect, the consolidation phase

begins. During this period, the new bone in the distraction gap bridges and corticalizes. The ring fixator is left in place during this time to allow the new bone to consolidate and become strong enough to withstand fracture, shortening, or bending. Historically, the length of this period has been described as two to three times the distraction period. Some authors have described an external fixator index (defined as the amount of time in months required per centimeter of distraction). Most report this index between 1.4 and 2.1 [97, 100, 101], although others have recently reported indices as low as 0.4 with the concomitant use of IM nails [32].

The Ilizarov technique has been reported with good results in all long bones in the body [102]. In a meta-analysis of lower limb segmental defects treated by bone transport, the overall union rate was 95% (range, 60–100%) [102]. Additionally, this technique can be used to correct alignments in any plane or direction including rotational deformities since it utilizes a circumferential ring fixator.

Although the Ilizarov technique is well established, disadvantages of the method include prolonged treatment times (up to years, depending on the length of the original defect), pin site infection (over 80% in some series [20, 47]), pin breakage, and the inconvenience and burden of prolonged circular external fixation [16, 97, 99, 102]. The soft tissues surrounding the defect are also manipulated and pulled, which can cause significant pain from muscle and nerve stretching, and can also cause neighboring joint contractures [98]. Finally, and critically, this technique requires significant cooperation from the patient, and patients must be informed of the time commitment involved in the procedure. Notably, although the amputation rate was only 2.6% in a meta-analysis of 37 studies of bone transport, half of these amputations were secondary to patient request [102]. This suggests that very careful explanation of the details of this technique and careful patient selection is important for this procedure.

Induced membrane technique

Recently, a new and promising technique has been increasingly utilized for the treatment of long bone segmental defects. In 2000, Masquelet first reported the results of this technique in a case series of 31 patients with segmental diaphyseal defects ranging from 5 to 25 cm [84]. This technique, which he termed “induced membrane,” is a two-stage procedure that utilizes a temporary cement spacer initially, followed by subsequent bone grafting [83]. The first stage consists of radical soft tissue and bone debridement, and implantation of a polymethylmethacrylate cement spacer at the bone defect. Stabilization is provided by a plate or nail. The cement spacer induces a foreign body reaction, which causes a fibrous membrane to form around the spacer.

During the second stage, which is typically performed 6–8 weeks later, the membrane is incised and the spacer is removed, and autograft is packed into the pseudocapsule. If the amount of autograft is insufficient, allograft is added to the mixture [41].

The cement spacer provides several mechanical and biological advantages. First, it prevents hematoma formation and fibrous tissue ingrowth into the bone defect and maintains the potential space as well as bone length and soft tissue tension. Secondly, the spacer can be impregnated with high dose antibiotics, and lack of infection after 6 weeks is an indication of favorable conditions for bone grafting. Finally, the fibrous membrane which forms around the spacer is highly vascularized and contains various osteoinductive factors including bone morphogenetic protein-2, vascularized endothelial growth factor, transcription growth factor-beta-1, and IL-6, IL-8, type I collagen, and von Willebrand factor [3, 25, 106]. Additionally, the membrane contains osteogenic properties: mesenchymal stem cells have been shown to be present in the membrane in both animal and human studies [25, 45, 106]. This vascularized pseudomembrane therefore provides support to the implanted autograft, inhibits graft resorption, and facilitates graft vascularization and corticalization/osseous consolidation [25, 106]. By contrast, studies have shown that membranes induced in subcutaneous or intramuscular sites do not contain mesenchymal stem cells and are less well vascularized [50, 73].

Since Masquelet's first report in 2000, several case series and retrospective studies utilizing this method have been published (Table 1). However, there are relatively few such studies and no direct comparisons of the induced membrane technique to other methods. Therefore, a consensus on the efficacy of this method versus other methods has yet to be determined. The technique has been successfully utilized in a variety of causes of critical-sized bone defects, including osteomyelitis, tumor resection, infected nonunions, and post-traumatic bone loss. Results are favorable, with a union rate of 88–100% in trauma cases, although lower rates have been reported after tumor resection (Table 1) [4, 44, 62, 82, 105, 117, 119, 122, 135, 138, 140]. Unfortunately, the methodology of the relevant studies varies considerably with regard to type of fixation (screws, plates, nails, external fixators) as well as type of graft (autologous or allogeneic and with or without the addition of BMP). For bone grafting, fresh cancellous autograft is the gold standard. However, large defects require large amounts of bone graft, and harvesting autologous bone is limited in quantity and carries morbidity [43, 64]. The ideal graft composition, as well as type of fixation, is yet to be determined.

Another question to be addressed is the optimal time between the first and second stages. Masquelet initially recommended 6–8 weeks [82, 84]. However, studies examining

the composition of the membrane have shown that the time of highest biological activity may be 4 weeks post-spacer placement [3, 50, 106]. For example, BMP-2 levels in the membrane peak at 4 weeks [106], and cells grown in culture from membranes between 2 and 4 weeks of age proliferated at a high rate while cells grown in culture from membranes taken at 6 weeks showed negligible proliferation [50]. The highest levels of vascularity and cell density were also found at 4 weeks [3, 45]. Based on these findings, some authors have advocated for earlier bone grafting. Several case series have reported successful union after grafting at 4 weeks [3, 136].

Reported complications of the induced membrane technique include infection, amputation, malunion, fracture, and nonunion necessitating reoperation and additional bone grafting. The rates of each of these complications is quite variable (see Table 1); however, infection is overall most common, ranging from 16.1% in Masquelet's original paper to 50% in a large study by Karger et al. of 84 patients [84]. Rates of reoperation range from 4 to 35% (Table 1). Karger et al. reported a mean number of 6.1 interventions from the first stage until union was obtained. Finally, cases of massive graft resorption have been reported in pediatric populations undergoing reconstruction after tumor resection [1, 22, 44]. In these patients, the second stage was not performed until after chemotherapy completion, 6 months after the initial stage, suggesting these failures could be due, at least in part, to delay and membrane devascularization. However, the considerable inhomogeneity present both within and between series in terms of defect cause, size, and location, as well as fracture complexity and associated comorbidities, precludes any definitive exploration of the rates of complications associated with the technique. Perhaps the most significant disadvantage of the Masquelet technique is the need for two surgeries. There has been a recent push toward a single-stage tissue engineered solution to surgically treat critical-sized bone loss.

Tissue engineering

The ideal tissue engineering construct for critical-sized bone defects would provide an environment that mimics the body's natural healing process. Such a graft would promote osteogenesis and angiogenesis, while having sufficient mechanical strength to promote integration with host tissues and facilitate load transfer under weight-bearing conditions. To date, no such construct has been developed. However, there are many promising techniques. This section will provide an up to date review of scaffold materials, growth factors, and methods of vascularization.

Scaffolds must be biocompatible, contain macro- and microporosity, have sufficient mechanical strength for load

Table 1 Comparison of studies utilizing the induced membranes technique including rate of union, time to union, average diaphyseal defect, and complications

Study	Study type	N	Indications	Defect size, mean (cm)	Union rate	Mean time to union, months	Infection	Other complications
Masquelet 2000 [79]	Prospective	31	Trauma	5–25	100%	4	16.1%	Refracture (12.9%), amputations (6.5%)
Schöttle 2005 [123]	Retrospective	6	Trauma, infected nonunion	6.5 (5–8)	83%	6.8	0%	Refracture (17%), thrombosis of free flap anastomosis (17%)
Ristinieni 2007 [124]	Retrospective	23	Trauma	5.2 (3.5–10)	96%	10		Reoperations for osseous healing (35%), 17 reoperations for soft tissue complications (74%)
Zwetyenga 2009 [125]	Case series	4	Osteoradionecrosis	11.3 (9–14)	50%	Not reported	50%	
Stafford 2010 [126]	Retrospective	27	Trauma nonunion	5.8 (1–25)	88%	Not reported	4%	BKA (25%), repeat grafting (25%)
Apard 2010 [89]	Retrospective	12	Trauma infected nonunion, aseptic nonunion	8.5 (4–15)	92%	Not reported.	33%	
Donegan 2010 [127]	Retrospective	11	Trauma, infected nonunion, aseptic nonunion, tumor	8.5 (4–15)	91%	7.5	9%	Heterotopic ossification (18%)
Zappaterra 2011 [98]	Case series	9	Trauma, infected nonunion, aseptic nonunion, tumor. Upper extremity.	5.9 (2.5–8)	100%	14.5		
Villemagne 2011 [96]	Retrospective	12	Pediatric tumor resection		58%	4.1	0%	Fractures (42%), femoral varus deformities (17%)
Sales de Gauzy 2012 [94]	Retrospective	10	Pediatric trauma, infected nonunion	3.5	100%	11.5	60%	Functional limitations (40%)
Karger 2012 [91]	Retrospective	84	Trauma, infected nonunion	6.8 (6.4–7.1)	91%	14.9	50%	Malalignment (14%), 6.1 mean procedures to achieve union
Chotel 2012 [102]	Prospective	8	Pediatric tumor resection	15 (10–22)	88%	4.8	0%	Paradoxical graft resorption (13%)
Gouron 2013 [90]	Retrospective review	14	Pediatric tumor resection, trauma, pseudarthrosis	10.3 (3.8–19.2)	100%	9.5	0%	Massive graft resorption (7%); reoperations 36% (autogenous grafting, better fixation)
Accadbled 2013 [101]	Case series	3	Pediatric tumor resection	19 (15–22)	0	N/A	0%	Massive graft resorptions (100%)
Scholz 2015 [95]	Case series	13	Septic diaphyseal defects	8 (5.5–14.5)	100%	4.2	0%	Malalignment requiring reoperation (8%), functional deficits (15%) (ROM, leg length difference)
Olesen 2015 [128]	Retrospective	8	Trauma, infected nonunion	5 (3–9)	75%	7	0%	Malalignment (25%), grafting needed (12%)
El-Alfy 2015 [129]	Retrospective	17	Infected nonunion, osteomyelitis	7 (4–11)	82%	10	12% reinfections	6% refracture

transfer, and also be bioresorbable. Biocompatibility refers to the ability to support cellular activity without toxicity to the host. The interconnected porosity is necessary for ingrowth and diffusion of nutrients; unfortunately, porosity reduces mechanical properties such as compressive strength. Additionally, cortical bone and cancellous bone vary significantly in their mechanical properties, which makes it difficult to design a single optimum scaffold.

Ceramics are a common material for bone scaffolds and include bioactive glass, tricalcium phosphate, and hydroxyapatite. All are biocompatible, osteoconductive, and have pores to allow tissue ingrowth. However, ceramics are brittle [115]. To overcome this shortcoming, synthetic biodegradable polymers such as polylactic acid (PLA), polyglycolic acid (PGA), or poly lactic acid-co-glycolic acid (PLGA) have been used to coat bioglass, hydroxyapatite, and beta-tricalcium phosphate, improving their mechanical properties as well as increasing their osteogenic potential [7, 8, 61]. These materials are bioresorbable synthetic polymers, which serve as scaffolds with good osteoinductive qualities. Moreover, although alone these compounds have poor mechanical properties for load-bearing, as well as poor osteoconductive properties, when combined with ceramics they have significantly increased strength [87, 118]. Combining PLA, PGA, or PLGA with the strength of ceramics is therefore a synergistic partnership.

Natural substitutes used for scaffolds include collagen, alginate, and hyaluronic acid. These each have excellent porosity and viscosity to allow for cellular immobilization and release of cells and osteoinductive factors from the scaffold; however, they lack sufficient mechanical strength [57, 69]. Similar to polymers, they can be combined with other compounds such as hydroxyapatite or chitosan to increase strength. One study found that a collagen–hydroxyapatite scaffold containing osteoprogenitor cells resulted in near complete filling of a 3.5-mm calvarial bone defect at 5 weeks, with much higher rates of scaffold breakdown compared to pure hydroxyapatite scaffolds [134]. Similarly encouraging results have been reported in other collagen–hydroxyapatite scaffolds [77, 109]. This resorbability is unique compared to pure hydroxyapatite scaffolds—for example, in one study, pure hydroxyapatite scaffolds used in critical-sized defects did not show any signs of resorption even after 7 years post-implantation [80]. This lack of resorption may create inferior mechanical qualities in the new bone, as well as represent an infection risk. Additionally, the strength of collagen scaffolds has recently been improved by novel modifications of collagen cross-linking [132].

Next-generation synthetics improve on the osteogenic properties of polymer–ceramic scaffolds through a variety of mechanisms. The addition of bone marrow mesenchymal stem cells and the incorporation of growth factors into the

scaffold have been shown to be associated with increased rate and quantity of bone formation compared to scaffolds without these additions [19, 74]. Additionally, the manner of release of growth factors is essential [60], and importantly, new technologies have allowed growth factors to be released from the scaffold continuously for up to 6 weeks, rather than all at once [58, 125].

Several studies using novel polymers to deliver growth factors have found increased bone formation at lower doses compared to the standard clinical delivery system for BMP-2 (collagen sponge). In one study, a polyethylene glycol synthetic polymer was used to encapsulate and deliver BMP-2 in a critical-sized defect in rat calvariums and showed significantly increased bone formation compared to collagen–sponge delivery systems [81]. In a sheep model of 3-cm tibial defects, implantation of synthetic scaffolds containing BMP-7 resulted in the generation of similar bone volume and torsional stiffness when compared to autologous bone graft [112]. Finally, the addition of integrin signaling has been shown to have exciting results: in scaffolds coated with GFOGER (which binds to cells via $\alpha 2 \beta 1$ integrin) as a BMP-2 delivery carrier, the subsequent bone formed was equal in torsional strength to that of native bone [124]. Moreover, this delivery system outperformed that of collagen sponges. Specifically, the GFOGER coated polymer stimulated bone healing at levels of BMP-2 that produced no results when delivered from collagen sponges.

Ideally, a graft would encourage angiogenesis around or even within the graft in order to perfuse the forming tissue. Methods of increasing vascularity around the scaffold include seeding scaffolds with endothelial progenitor cells and angiogenic agents such as VEGF. Such seeding is associated with increased vascularization compared to non-seeded scaffolds and promotes increased bone healing when coupled with the addition of mesenchymal stem cells [123, 142].

A tissue engineered vascularized bone graft might someday achieve the ideal graft characteristics through the incorporation of cell-laden hydrogels and synthetic vascular grafts into a porous osteoconductive rigid frame. The soft hydrogel would be utilized to control the spatial and temporal distribution of cells and growth factors, while the incorporation of vessel graft beds would allow for immediate perfusion. A rigid channeled macroporous frame with structural and mechanical properties resembling those of bone would promote integration and facilitate load transfer [88].

Given the difficulty of engineering functional tissue that recapitulates the composition, biomechanical properties, and physiological performance of native tissues, *in vivo* bioreactors utilize the body to grow grafts for later transplantation to the defect site. A scaffold within a chamber can be implanted into the body, normally intramuscularly or adjacent to periosteum and the osteogenic and angiogenic

capacity of the body is exploited. With time, a vascularized bone graft is generated. The implanted chamber and scaffold can be modified to the desired geometry and harvested with vessels for the treatment of bone defects [86, 131].

Conclusions

Critical-sized bone defects remain one of the most challenging orthopedic conditions to treat. Several techniques have been described, each with their own unique advantages and disadvantages. Distraction osteogenesis and free fibular grafting have been the historical mainstays of treatment. Although good clinical results have been reported for both methods, distraction osteogenesis requires lengthy treatment times and is associated with a high incidence of pin site infections, while fibular grafts are highly technically demanding and may not be ideal for lower extremity defects. The more recent induced membrane technique is an increasingly utilized method with favorable results; however, it does require at least two stages and is not as predictable as would be hoped. Tissue engineering holds great promise in the treatment of large bone defects due to advancement of stem cell biology, novel biomaterials, and 3D bioprinting, but the clinical implementation will be dependent upon whether the outcome and efficacy of this approach can provide additional benefits compared to the current clinical treatments, as well as upon the collaborative movement on commercialization, quality and safety control, and regulatory issues.

Acknowledgements Generous support from Kent Thiry and Denise O’Leary, Boswell Foundation, NIH R01AR057837 (NIAMS), and NIH 1U01AR069395 (NIAMS).

Compliance with ethical standards

Conflict of interest Drs. Roddy, DeBaun, Daoud, and Yang have no conflicts of interest to declare. Dr. Gardner reports personal fees from DePuy-Synthes, personal fees from KCI, personal fees from Miami Medical, personal fees from Biocomposites, personal fees from Pacira, outside the submitted work.

References

- Accadbled F, Mazeau P, Chotel F, Cottalorda J, Sales de Gauzy J, Kohler R (2013) Induced-membrane femur reconstruction after resection of bone malignancies: three cases of massive graft resorption in children. *Orthop Traumatol Surg Res OTSR* 99:479–483. doi:10.1016/j.otsr.2013.01.008
- Ackermann PW, Hart DA (2013) Influence of comorbidities: neuropathy, vasculopathy, and diabetes on healing response quality. *Adv Wound Care* 2:410–421. doi:10.1089/wound.2012.0437
- Aho OM, Lehenkari P, Ristiniemi J, Lehtonen S, Risteli J, Leskela HV (2013) The mechanism of action of induced membranes in bone repair. *J Bone Jt Surg Am* 95:597–604. doi:10.2106/JBJS.L.00310
- Apard T, Bigorre N, Cronier P, Duteille F, Bizot P, Massin P (2010) Two-stage reconstruction of post-traumatic segmental tibia bone loss with nailing. *Orthop Traumatol Surg Res OTSR* 96:549–553. doi:10.1016/j.otsr.2010.02.010
- Aponte-Tinao L, Farfalli GL, Ritacco LE, Ayerza MA, Muscolo DL (2012) Intercalary femur allografts are an acceptable alternative after tumor resection. *Clin Orthop Relat Res* 470:728–734. doi:10.1007/s11999-011-1952-5
- Arai K, Toh S, Tsubo K, Nishikawa S, Narita S, Miura H (2002) Complications of vascularized fibula graft for reconstruction of long bones. *Plast Reconstr Surg* 109:2301–2306
- Ardeshirylajimi A et al (2015) Enhanced osteoconductivity of polyethersulphone nanofibres loaded with bioactive glass nanoparticles in in vitro and in vivo models. *Cell Prolif* 48:455–464. doi:10.1111/cpr.12198
- Ardeshirylajimi A, Hosseinkhani S, Parivar K, Yaghmaie P, Soleimani M (2013) Nanofiber-based polyethersulfone scaffold and efficient differentiation of human induced pluripotent stem cells into osteoblastic lineage. *Mol Biol Rep* 40:4287–4294. doi:10.1007/s11033-013-2515-5
- Aronson J (1997) Limb-lengthening, skeletal reconstruction, and bone transport with the Ilizarov method. *J Bone Jt Surg Am* 79:1243–1258
- Aronson J, Harrison B, Boyd CM, Cannon DJ, Lubansky HJ (1988) Mechanical induction of osteogenesis: the importance of pin rigidity. *J Pediatr Orthop* 8:396–401
- Aronson J, Harrison B, Boyd CM, Cannon DJ, Lubansky HJ, Stewart C (1988) Mechanical induction of osteogenesis. *Prelim Stud Ann Clin Lab Sci* 18:195–203
- Aronson J, Johnson E, Harp JH (1989) Local bone transportation for treatment of intercalary defects by the Ilizarov technique. Biomechanical and clinical considerations. *Clin Orthop Relat Res* 243:71–79
- Bauer TW, Muschler GF (2000) Bone graft materials. An overview of the basic science. *Clin Orthop Relat Res* 371:10–27
- Beris AE, Lykissas MG, Korompilias AV, Vekris MD, Mitsionis GI, Malizos KN, Soucacos PN (2011) Vascularized fibula transfer for lower limb reconstruction. *Microsurgery* 31:205–211. doi:10.1002/micr.20841
- Bilic R et al (2006) Osteogenic protein-1 (BMP-7) accelerates healing of scaphoid non-union with proximal pole sclerosis. *Int Orthop* 30:128–134. doi:10.1007/s00264-005-0045-z
- Blum AL, BongioVanni JC, Morgan SJ, Flierl MA, dos Reis FB (2010) Complications associated with distraction osteogenesis for infected nonunion of the femoral shaft in the presence of a bone defect: a retrospective series. *J Bone Jt Surg Br* 92:565–570. doi:10.1302/0301-620X.92B4.23475
- Bullens PH, Minderhoud NM, de Waal Malefijt MC, Veth RP, Buma P, Schreuder HW (2009) Survival of massive allografts in segmental oncological bone defect reconstructions. *Int Orthop* 33:757–760. doi:10.1007/s00264-008-0700-2
- Ceruso M, Taddei F, Bigazzi P, Manfrini M (2008) Vascularised fibula graft inlaid in a massive bone allograft: considerations on the bio-mechanical behaviour of the combined graft in segmental bone reconstructions after sarcoma resection. *Injury* 39(Suppl 3):S68–74. doi:10.1016/j.injury.2008.05.014
- Cha JK, Lee JS, Kim MS, Choi SH, Cho KS, Jung UW (2014) Sinus augmentation using BMP-2 in a bovine hydroxyapatite/collagen carrier in dogs. *J Clin Periodontol* 41:86–93. doi:10.1111/jcpe.12174
- Chaddha M, Gulati D, Singh AP, Singh AP, Maini L (2010) Management of massive posttraumatic bone defects in the lower limb with the Ilizarov technique. *Acta Orthop Belg* 76:811–820
- Chakkalakal DA (2005) Alcohol-induced bone loss and deficient bone repair. *Alcohol Clin Exp Res* 29:2077–2090

22. Chotel F, Nguiabanda L, Braillon P, Kohler R, Berard J, Abelin-Genevois K (2012) Induced membrane technique for reconstruction after bone tumor resection in children: a preliminary study. *Orthop Traumatol Surg Res OTSR* 98:301–308. doi:[10.1016/j.otsr.2011.11.008](https://doi.org/10.1016/j.otsr.2011.11.008)
23. Claes L, Recknagel S, Ignatius A (2012) Fracture healing under healthy and inflammatory conditions Nature reviews. *Rheumatology* 8:133–143. doi:[10.1038/nrrheum.2012.1](https://doi.org/10.1038/nrrheum.2012.1)
24. Conway JD (2010) Autograft and nonunions: morbidity with intramedullary bone graft versus iliac crest bone graft. *Orthop Clin N Am* 41:75–84. doi:[10.1016/j.ocl.2009.07.006](https://doi.org/10.1016/j.ocl.2009.07.006) (table of contents)
25. Cuthbert RJ, Churchman SM, Tan HB, McGonagle D, Jones E, Giannoudis PV (2013) Induced periosteum a complex cellular scaffold for the treatment of large bone defects. *Bone* 57:484–492. doi:[10.1016/j.bone.2013.08.009](https://doi.org/10.1016/j.bone.2013.08.009)
26. Daftari TK, Whitesides TE Jr, Heller JG, Goodrich AC, McCarey BE, Hutton WC (1994) Nicotine on the revascularization of bone graft. *Exp Study Rabbits Spine* 19:904–911
27. de Boer HH, Wood MB (1989) Bone changes in the vascularised fibular graft. *J Bone Jt Surg Br* 71:374–378
28. de Boer HH, Wood MB, Hermans J (1990) Reconstruction of large skeletal defects by vascularized fibula transfer. Factors that influenced the outcome of union in 62 cases. *Int Orthop* 14:121–128
29. Dimitriou R, Mataliotakis GI, Angoules AG, Kanakaris NK, Giannoudis PV (2011) Complications following autologous bone graft harvesting from the iliac crest and using the RIA: a systematic review. *Injury* 42(Suppl 2):S3–15. doi:[10.1016/j.injury.2011.06.015](https://doi.org/10.1016/j.injury.2011.06.015)
30. Dimitriou R, Tsiridis E, Giannoudis PV (2005) Current concepts of molecular aspects of bone healing. *Injury* 36:1392–1404. doi:[10.1016/j.injury.2005.07.019](https://doi.org/10.1016/j.injury.2005.07.019)
31. Emara KM, Ghafar KA, Al Kersh MA (2011) Methods to shorten the duration of an external fixator in the management of tibial infections. *World J Orthop* 2:85–92. doi:[10.5312/wjo.v2.i9.85](https://doi.org/10.5312/wjo.v2.i9.85)
32. Eward WC, Kontogeorgakos V, Levin LS, Brigman BE (2010) Free vascularized fibular graft reconstruction of large skeletal defects after tumor resection. *Clin Orthop Relat Res* 468:590–598. doi:[10.1007/s11999-009-1053-x](https://doi.org/10.1007/s11999-009-1053-x)
33. Farfalli GL, Aponte-Tinao L, Lopez-Millan L, Ayerza MA, Muscolo DL (2012) Clinical and functional outcomes of tibial intercalary allografts after tumor resection. *Orthopedics* 35:e391–396. doi:[10.3928/01477447-20120222-25](https://doi.org/10.3928/01477447-20120222-25)
34. Foulk DA, Szabo RM (1995) Diaphyseal humerus fractures: natural history and occurrence of nonunion. *Orthopedics* 18:333–335
35. Fourman MS, Borst EW, Bogner E, Rozbruch SR, Fragomen AT (2014) Recombinant human BMP-2 increases the incidence and rate of healing in complex ankle arthrodesis. *Clin Orthop Relat Res* 472:732–739. doi:[10.1007/s11999-013-3261-7](https://doi.org/10.1007/s11999-013-3261-7)
36. Friedlaender GE et al (2001) Osteogenic protein-1 (bone morphogenetic protein-7) in the treatment of tibial nonunions. *J Bone Jt Surg Am* 83-A(Suppl 1):S151–S158
37. Friedrich JB, Moran SL, Bishop AT, Wood CM, Shin AY (2008) Free vascularized fibular graft salvage of complications of long-bone allograft after tumor reconstruction. *J Bone Jt Surg Am* 90:93–100. doi:[10.2106/JBJS.G.00551](https://doi.org/10.2106/JBJS.G.00551)
38. Garrison KR et al (2010) Bone morphogenetic protein (BMP) for fracture healing in adults. *Cochrane Database Syst Rev*. doi:[10.1002/14651858.CD006950.pub2](https://doi.org/10.1002/14651858.CD006950.pub2)
39. Gaston MS, Simpson AH (2007) Inhibition of fracture healing. *J Bone Jt Surg Br* 89:1553–1560. doi:[10.1302/0301-620X.89B12.19671](https://doi.org/10.1302/0301-620X.89B12.19671)
40. Giannoudis PV, Dinopoulos H, Tsiridis E (2005) Bone substitutes: an update. *Injury* 36(Suppl 3):S20–27. doi:[10.1016/j.injury.2005.07.029](https://doi.org/10.1016/j.injury.2005.07.029)
41. Giannoudis PV, Faour O, Goff T, Kanakaris N, Dimitriou R (2011) Masquelet technique for the treatment of bone defects: tips-tricks and future directions. *Injury* 42:591–598. doi:[10.1016/j.injury.2011.03.036](https://doi.org/10.1016/j.injury.2011.03.036)
42. Goel A, Sangwan SS, Siwach RC, Ali AM (2005) Percutaneous bone marrow grafting for the treatment of tibial non-union. *Injury* 36:203–206. doi:[10.1016/j.injury.2004.01.009](https://doi.org/10.1016/j.injury.2004.01.009)
43. Goulet JA, Senunas LE, DeSilva GL, Greenfield ML (1997) Autogenous iliac crest bone graft. Complications and functional assessment. *Clin Orthop Relat Res* 339:76–81
44. Gouron R, Deroussen F, Plancq MC, Collet LM (2013) Bone defect reconstruction in children using the induced membrane technique: a series of 14 cases. *Orthop Traumatol Surg Res OTSR* 99:837–843. doi:[10.1016/j.otsr.2013.05.005](https://doi.org/10.1016/j.otsr.2013.05.005)
45. Gouron R, Petit L, Boudot C, Six I, Brazier M, Kamel S, Mentaverri R (2014) Osteoclasts and their precursors are present in the induced-membrane during bone reconstruction using the Masquelet technique. *J Tissue Eng Regen Med*. doi:[10.1002/term.1921](https://doi.org/10.1002/term.1921)
46. Govender S et al (2002) Recombinant human bone morphogenetic protein-2 for treatment of open tibial fractures: a prospective, controlled, randomized study of four hundred and fifty patients. *J Bone Jt Surg Am* 84-A:2123–2134
47. Green SA (1994) Skeletal defects. A comparison of bone grafting and bone transport for segmental skeletal defects. *Clin Orthop Relat Res* 301:111–117
48. Haverstock BD, Mandracchia VJ (1998) Cigarette smoking and bone healing: implications in foot and ankle surgery. *J Foot Ankle Surg* 37:69–74 (discussion 78)
49. Heitmann C, Erdmann D, Levin LS (2002) Treatment of segmental defects of the humerus with an osteoseptocutaneous fibular transplant. *J Bone Jt Surg Am* 84-A:2216–2223
50. Henrich D et al (2013) Establishment and characterization of the Masquelet induced membrane technique in a rat femur critical-sized defect model. *J Tissue Eng Regen Med*. doi:[10.1002/term.1826](https://doi.org/10.1002/term.1826)
51. Hernandez RK, Do TP, Critchlow CW, Dent RE, Jick SS (2012) Patient-related risk factors for fracture-healing complications in the United Kingdom general practice research database. *Acta Orthop* 83:653–660. doi:[10.3109/17453674.2012.747054](https://doi.org/10.3109/17453674.2012.747054)
52. Hernigou P, Mathieu G, Pognard A, Manicou O, Beaujean F, Rouard H (2006) Percutaneous autologous bone-marrow grafting for nonunions. Surgical technique. *J Bone Jt Surg Am* 88(Suppl 1 Pt 2):322–327. doi:[10.2106/JBJS.F.00203](https://doi.org/10.2106/JBJS.F.00203)
53. Higgins TF, Casey V, Bachus K (2007) Cortical heat generation using an irrigating/aspirating single-pass reaming vs conventional stepwise reaming. *J Orthop Trauma* 21:192–197. doi:[10.1097/BOT.0b013e318038d952](https://doi.org/10.1097/BOT.0b013e318038d952)
54. Houdek MT, Wagner ER, Stans AA, Shin AY, Bishop AT, Sim FH, Moran SL (2016) What is the outcome of allograft and intramedullary free fibula (capanna technique) in pediatric and adolescent patients with bone tumors? *Clin Orthop Relat Res* 474:660–668. doi:[10.1007/s11999-015-4204-2](https://doi.org/10.1007/s11999-015-4204-2)
55. Huddleston PM, Steckelberg JM, Hanssen AD, Rouse MS, Bolander ME, Patel R (2000) Ciprofloxacin inhibition of experimental fracture healing. *J Bone Jt Surg Am* 82:161–173
56. Ja K (1934) The effect of a local calcium depot on osteogenesis and healing of fractures. *J Bone Jt Surg Am* 16:176–184
57. Jin HH, Kim DH, Kim TW, Shin KK, Jung JS, Park HC, Yoon SY (2012) In vivo evaluation of porous hydroxyapatite/chitosan-alginate composite scaffolds for bone tissue engineering. *Int J Biol Macromol* 51:1079–1085. doi:[10.1016/j.ijbiomac.2012.08.027](https://doi.org/10.1016/j.ijbiomac.2012.08.027)

58. Jun SH, Lee EJ, Jang TS, Kim HE, Jang JH, Koh YH (2013) Bone morphogenic protein-2 (BMP-2) loaded hybrid coating on porous hydroxyapatite scaffolds for bone tissue engineering. *J Mater Sci Mater Med* 24:773–782. doi:[10.1007/s10856-012-4822-0](https://doi.org/10.1007/s10856-012-4822-0)
59. Kanakaris NK et al (2009) Application of bone morphogenetic proteins to femoral non-unions: a 4-year multicentre experience. *Injury* 40(Suppl 3):S54–S61. doi:[10.1016/S0020-1383\(09\)70013-0](https://doi.org/10.1016/S0020-1383(09)70013-0)
60. Kaneda K, Kurakami C, Minami A (1988) Free vascularized fibular strut graft in the treatment of kyphosis. *Spine* 13:1273–1277
61. Kang Y, Scully A, Young DA, Kim S, Tsao H, Sen M, Yang Y (2011) Enhanced mechanical performance and biological evaluation of a PLGA coated beta-TCP composite scaffold for load-bearing applications. *Eur Polym J* 47:1569–1577. doi:[10.1016/j.eurpolymj.2011.05.004](https://doi.org/10.1016/j.eurpolymj.2011.05.004)
62. Karger C, Kishi T, Schneider L, Fitoussi F, Masquelet AC, French Society of Orthopaedic S, Traumatology (2012) Treatment of posttraumatic bone defects by the induced membrane technique. *Orthop Traumatol Surg Res OTSR* 98:97–102. doi:[10.1016/j.otsr.2011.11.001](https://doi.org/10.1016/j.otsr.2011.11.001)
63. Keating JF, Simpson AH, Robinson CM (2005) The management of fractures with bone loss. *J Bone Jt Surg Br* 87:142–150
64. Kim DH et al (2009) Prospective study of iliac crest bone graft harvest site pain and morbidity. *Spine J* 9:886–892. doi:[10.1016/j.spinee.2009.05.006](https://doi.org/10.1016/j.spinee.2009.05.006)
65. Kline AJ, Gruen GS, Pape HC, Tarkin IS, Irrgang JJ, Wukich DK (2009) Early complications following the operative treatment of pilon fractures with and without diabetes. *Foot Ankle Int* 30:1042–1047. doi:[10.3113/FAI.2009.1042](https://doi.org/10.3113/FAI.2009.1042)
66. Kyro A, Usenius JP, Aarnio M, Kunnamo I, Avikainen V (1993) Are smokers a risk group for delayed healing of tibial shaft fractures? *Ann Chir Gynaecol* 82:254–262
67. Lee KS, Han SB, Baek JR (2004) Free vascularized osteocutaneous fibular graft to the tibia in 51 consecutive cases. *J Reconstr Microsurg* 20:277–284. doi:[10.1055/s-2004-824884](https://doi.org/10.1055/s-2004-824884)
68. Li J, Wang Z, Guo Z, Chen GJ, Fu J, Pei GX (2010) The use of allograft shell with intramedullary vascularized fibula graft for intercalary reconstruction after diaphyseal resection for lower extremity bony malignancy. *J Surg Oncol* 102:368–374. doi:[10.1002/jso.21620](https://doi.org/10.1002/jso.21620)
69. Li Z, Ramay HR, Hauch KD, Xiao D, Zhang M (2005) Chitosan-alginate hybrid scaffolds for bone tissue engineering. *Biomaterials* 26:3919–3928. doi:[10.1016/j.biomaterials.2004.09.062](https://doi.org/10.1016/j.biomaterials.2004.09.062)
70. Liang K, Xiang Z, Chen S, Cen S, Zhong G, Yi M, Huang F (2012) Folded free vascularized fibular grafts for the treatment of subtrochanteric fractures complicated with segmental bone defects. *J Trauma Acute Care Surg* 72:1404–1410. doi:[10.1097/TA.0b013e31824473ce](https://doi.org/10.1097/TA.0b013e31824473ce)
71. Lin CH, Wei FC, Chen HC, Chuang DC (1999) Outcome comparison in traumatic lower-extremity reconstruction by using various composite vascularized bone transplantation. *Plast Reconstr Surg* 104:984–992
72. Ling XF, Peng X (2012) What is the price to pay for a free fibula flap? A systematic review of donor-site morbidity following free fibula flap surgery. *Plast Reconstr Surg* 129:657–674. doi:[10.1097/PRS.0b013e3182402d9a](https://doi.org/10.1097/PRS.0b013e3182402d9a)
73. Liu H, Hu G, Shang P, Shen Y, Nie P, Peng L, Xu H (2013) Histological characteristics of induced membranes in subcutaneous, intramuscular sites and bone defect. *Orthop Traumatol Surg Res OTSR* 99:959–964. doi:[10.1016/j.otsr.2013.08.009](https://doi.org/10.1016/j.otsr.2013.08.009)
74. Liu Y, Ming L, Luo H, Liu W, Zhang Y, Liu H, Jin Y (2013) Integration of a calcined bovine bone and BMSC-sheet 3D scaffold and the promotion of bone regeneration in large defects. *Biomaterials* 34:9998–10006. doi:[10.1016/j.biomaterials.2013.09.040](https://doi.org/10.1016/j.biomaterials.2013.09.040)
75. Loder RT (1988) The influence of diabetes mellitus on the healing of closed fractures. *Clin Orthop Relat Res* 232:210–216
76. Lowe JA, Della Rocca GJ, Murtha Y, Liporace FA, Stover MD, Nork SE, Crist BD (2010) Complications associated with negative pressure reaming for harvesting autologous bone graft: a case series. *J Orthop Trauma* 24:46–52. doi:[10.1097/BOT.0b013e31819c0ccb](https://doi.org/10.1097/BOT.0b013e31819c0ccb)
77. Lyons FG, Gleeson JP, Partap S, Coghlan K, O'Brien FJ (2014) Novel microhydroxyapatite particles in a collagen scaffold: a bioactive bone void filler? *Clin Orthop Relat Res* 472(4):1318–1328
78. Maeda M, Bryant MH, Yamagata M, Li G, Earle JD, Chao EY (1988) Effects of irradiation on cortical bone and their time-related changes. A biomechanical and histomorphological study. *J Bone Jt Surg Am* 70:392–399
79. Malizos KN, Zalavras CG, Soucacos PN, Beris AE, Urbaniak JR (2004) Free vascularized fibular grafts for reconstruction of skeletal defects. *J Am Acad Orthop Surg* 12:360–369
80. Marcacci M, Kon E, Moukhachev V, Lavroukov A, Kutepov S, Quarto R, Mastrogiacomo M, Cancedda R (2007) Stem cells associated with macroporous bioceramics for long bone repair: 6- to 7-year outcome of a pilot clinical study. *Tissue Eng* 13(5):947–955
81. Mariner PD, Wudel JM, Miller DE, Genova EE, Streubel SO, Anseth KS (2013) Synthetic hydrogel scaffold is an effective vehicle for delivery of INFUSE (rhBMP2) to critical-sized calvaria bone defects in rats. *J Orthop Res* 31:401–406. doi:[10.1002/jor.22243](https://doi.org/10.1002/jor.22243)
82. Masquelet AC (2003) Muscle reconstruction in reconstructive surgery: soft tissue repair and long bone reconstruction *Langenbeck's archives of surgery / Deutsche Gesellschaft fur Chirurgie* 388:344–346. doi:[10.1007/s00423-003-0379-1](https://doi.org/10.1007/s00423-003-0379-1)
83. Masquelet AC, Begue T (2010) The concept of induced membrane for reconstruction of long bone defects. *Orthop Clin N Am* 41:27–37. doi:[10.1016/j.ocl.2009.07.011](https://doi.org/10.1016/j.ocl.2009.07.011) (table of contents)
84. Masquelet AC, Fitoussi F, Begue T, Muller GP (2000) Reconstruction of the long bones by the induced membrane and spongy autograft. *Ann Chir Plast Esthet* 45:346–353
85. McBride JCM, Banks RE, Taylor D, Ryan J (1993) Healing of segmental bone defects in goat tibia. *J Invest Surg* 6:369
86. McCullen SD, Chow AG, Stevens MM (2011) In vivo tissue engineering of musculoskeletal tissues. *Curr Opin Biotechnol* 22:715–720. doi:[10.1016/j.copbio.2011.05.001](https://doi.org/10.1016/j.copbio.2011.05.001)
87. Mercado-Pagan AE, Kang Y, Ker DF, Park S, Yao J, Bishop J, Yang Y (2013) Synthesis and characterization of novel elastomeric poly(D, L-lactide urethane) maleate composites for bone tissue engineering. *Eur Polym J* 49:3337–3349. doi:[10.1016/j.eurpolymj.2013.07.004](https://doi.org/10.1016/j.eurpolymj.2013.07.004)
88. Mercado-Pagan AE, Stahl AM, Shanjani Y, Yang Y (2015) Vascularization in bone tissue engineering constructs. *Ann Biomed Eng* 43:718–729. doi:[10.1007/s10439-015-1253-3](https://doi.org/10.1007/s10439-015-1253-3)
89. Minami A, Kasashima T, Iwasaki N, Kato H, Kaneda K (2000) Vascularized fibular grafts. An experience of 102 patients. *J Bone Jt Surg Br* 82:1022–1025
90. Muramatsu K, Ihara K, Shigetomi M, Kawai S (2004) Femoral reconstruction by single, folded or double free vascularised fibular grafts. *Br J Plast Surg* 57:550–555. doi:[10.1016/j.bjps.2003.08.021](https://doi.org/10.1016/j.bjps.2003.08.021)
91. Muscolo DL (2012) Accurate 3-dimensional preoperative planning and resection in orthopedic oncology. *Orthopedics* 35:7–8. doi:[10.3928/01477447-20111122-01](https://doi.org/10.3928/01477447-20111122-01)
92. Muscolo DL, Ayerza MA, Aponte-Tinao LA (2006) Massive allograft use in orthopedic oncology. *Orthop Clin N Am* 37:65–74. doi:[10.1016/j.ocl.2005.08.003](https://doi.org/10.1016/j.ocl.2005.08.003)
93. Ng VY (2012) Risk of disease transmission with bone allograft. *Orthopedics* 35:679–681. doi:[10.3928/01477447-20120725-04](https://doi.org/10.3928/01477447-20120725-04)
94. Noaman HH (2013) Management of upper limb bone defects using free vascularized osteoseptocutaneous fibular bone graft. *Ann Plast Surg* 71:503–509. doi:[10.1097/SAP.0b013e3182a1aff0](https://doi.org/10.1097/SAP.0b013e3182a1aff0)

95. Nusbickel FR, Dell PC, McAndrew MP, Moore MM (1989) Vascularized autografts for reconstruction of skeletal defects following lower extremity trauma. *Rev Clin Orthop Relat Res* 243:65–70
96. Ozaksar K, Sugun TS, Toros T, Gurbuz Y, Kayalar M, Ozerkan F (2012) Free vascularized fibular grafts in type 3 open tibia fractures. *Acta orthopaedica et traumatologica turcica* 46:430–437
97. Palatnik Y, Rozbruch SR (2011) Femoral reconstruction using external fixation. *Adv Orthop* 2011:967186. doi:10.4061/2011/967186
98. Paley D (1990) Problems, obstacles, and complications of limb lengthening by the Ilizarov technique. *Clin Orthop Relat Res* 250:81–104
99. Paley D, Catagni M, Argnani F, Prevot J, Bell D, Armstrong P (1992) Treatment of congenital pseudoarthrosis of the tibia using the Ilizarov technique. *Clin Orthop Relat Res* 250:81–93
100. Paley D, Herzenberg JE, Paremian G, Bhave A (1997) Femoral lengthening over an intramedullary nail. A matched-case comparison with Ilizarov femoral lengthening. *J Bone Jt Surg Am* 79:1464–1480
101. Paley D, Maar DC (2000) Ilizarov bone transport treatment for tibial defects. *J Orthop Trauma* 14:76–85
102. Papakostidis C, Bhandari M, Giannoudis PV (2013) Distraction osteogenesis in the treatment of long bone defects of the lower limbs: effectiveness, complications and clinical results; a systematic review and meta-analysis. *Bone Jt J* 95-B:1673–1680. doi:10.1302/0301-620X.95B12.32385
103. Pape HC, Zelle BA, Hildebrand F, Giannoudis PV, Krettek C, van Griensven M (2005) Reamed femoral nailing in sheep: does irrigation and aspiration of intramedullary contents alter the systemic response? *J Bone Jt Surg Am* 87:2515–2522. doi:10.2106/JBJS.D.02024
104. Patel RA, Wilson RF, Patel PA, Palmer RM (2013) The effect of smoking on bone healing: A systematic review. *Bone Jt Res* 2:102–111. doi:10.1302/2046-3758.26.2000142
105. Pelissier P, Bollecker V, Martin D, Baudet J (2002) Foot reconstruction with the “bi-Masquelet” procedure. *Ann Chir Plast Esthet* 47:304–307
106. Pelissier P, Masquelet AC, Bareille R, Pelissier SM, Amedee J (2004) Induced membranes secrete growth factors including vascular and osteoinductive factors and could stimulate bone regeneration. *J Orthop Res* 22:73–79. doi:10.1016/S0736-0266(03)00165-7
107. Porter SE, Hanley EN Jr (2001) The musculoskeletal effects of smoking. *J Am Acad Orthop Surg* 9:9–17
108. Qi Y, Sun HT, Fan YG, Li FM, Lin ZS (2016) Do stress fractures induce hypertrophy of the grafted fibula? A report of three cases received free vascularized fibular graft treatment for tibial defects. *Chin J Traumatol = Zhonghua chuang shang za zhi / Chin Med Assoc* 19:179–181
109. Quinlan E, Thompson EM, Matsiko A, O’Brien FJ, Lopez-Noriega A (2015) Long-term controlled delivery of rhBMP-2 from collagen-hydroxyapatite scaffolds for superior bone tissue regeneration. *J Control Release* 207:112–119
110. Qvick LM, Ritter CA, Mutty CE, Rohrbacher BJ, Buyea CM, Anders MJ (2013) Donor site morbidity with reamer-irrigator-aspirator (RIA) use for autogenous bone graft harvesting in a single centre 204 case series. *Injury* 44:1263–1269. doi:10.1016/j.injury.2013.06.008
111. Rabitsch K, Maurer-Ertl W, Pirker-Fruhauf U, Wibmer C, Leithner A (2013) Intercalary reconstructions with vascularised fibula and allograft after tumour resection in the lower limb. *Sarcoma* 2013:160295. doi:10.1155/2013/160295
112. Reichert JC et al (2012) A tissue engineering solution for segmental defect regeneration in load-bearing long bones. *Sci Transl Med* 4:141ra193. doi:10.1126/scitranslmed.3003720
113. Repo JP, Sommarhem A, Roine RP, Sintonen H, Halonen T, Tukiainen E (2016) Free vascularized fibular graft is reliable in upper extremity long-bone reconstruction with good long-term outcomes. *J Reconstr Microsurg*. doi:10.1055/s-0036-1581075
114. Retzeppi M, Donos N (2010) The effect of diabetes mellitus on osseous healing. *Clin Oral Implant Res* 21:673–681. doi:10.1111/j.1600-0501.2010.01923.x
115. Rezwani K, Chen QZ, Blaker JJ, Boccaccini AR (2006) Biodegradable and bioactive porous polymer/inorganic composite scaffolds for bone tissue engineering. *Biomaterials* 27:3413–3431. doi:10.1016/j.biomaterials.2006.01.039
116. Riebel GD, Boden SD, Whitesides TE, Hutton WC (1995) The effect of nicotine on incorporation of cancellous bone graft in an animal model. *Spine* 20:2198–2202
117. Ronga M, Ferraro S, Fagetti A, Cherubino M, Valdatta L, Cherubino P (2014) Masquelet technique for the treatment of a severe acute tibial bone loss. *Injury* 45(Suppl 6):S111–115. doi:10.1016/j.injury.2014.10.033
118. Roohani-Esfahani SI, Nouri-Khorasani S, Lu Z, Appleyard R, Zreiqat H (2010) The influence hydroxyapatite nanoparticle shape and size on the properties of biphasic calcium phosphate scaffolds coated with hydroxyapatite-PCL composites. *Biomaterials* 31:5498–5509. doi:10.1016/j.biomaterials.2010.03.058
119. Sales de Gauzy J et al (2012) Traumatic diaphyseal bone defects in children. *Orthop Traumatol Surg Res OTSR* 98:220–226. doi:10.1016/j.otsr.2012.01.001
120. Schmidmaier G, Herrmann S, Green J, Weber T, Scharfenberger A, Haas NP, Wildemann B (2006) Quantitative assessment of growth factors in reaming aspirate, iliac crest, and platelet preparation. *Bone* 39:1156–1163. doi:10.1016/j.bone.2006.05.023
121. Schmitz JP, Hollinger JO (1986) The critical size defect as an experimental model for craniomandibulofacial nonunions. *Clin Orthop Relat Res* 205:299–308
122. Scholz AO, Gehrman S, Glombitza M, Kaufmann RA, Bostelmann R, Flohe S, Windolf J (2015) Reconstruction of septic diaphyseal bone defects with the induced membrane technique. *Injury* 46(Suppl 4):S121–124. doi:10.1016/S0020-1383(15)30030-9
123. Seebach C, Henrich D, Kahling C, Wilhelm K, Tami AE, Alini M, Marzi I (2010) Endothelial progenitor cells and mesenchymal stem cells seeded onto beta-TCP granules enhance early vascularization and bone healing in a critical-sized bone defect in rats. *Tissue Eng Part A* 16:1961–1970. doi:10.1089/ten.TEA.2009.0715
124. Shekaran A, Garcia JR, Clark AY, Kavanaugh TE, Lin AS, Guldberg RE, Garcia AJ (2014) Bone regeneration using an alpha 2 beta 1 integrin-specific hydrogel as a BMP-2 delivery vehicle. *Biomaterials* 35:5453–5461. doi:10.1016/j.biomaterials.2014.03.055
125. Shi Y et al (2016) Evaluation of a novel HA/ZrO₂-based porous bioceramic artificial vertebral body combined with a rhBMP-2/chitosan slow-release hydrogel. *PLoS ONE* 11:e0157698. doi:10.1371/journal.pone.0157698
126. Song HR, Kale A, Park HB, Koo KH, Chae DJ, Oh CW, Chung DW (2003) Comparison of internal bone transport and vascularized fibular grafting for femoral bone defects. *J Orthop Trauma* 17:203–211
127. Soucacos PN, Kokkalis ZT, Piagkou M, Johnson EO (2013) Vascularized bone grafts for the management of skeletal defects in orthopaedic trauma and reconstructive surgery. *Injury* 44(Suppl 1):S70–75. doi:10.1016/S0020-1383(13)70016-0
128. Soucacos PN, Korompilias AV, Vekris MD, Zoubos A, Beris AE (2011) The free vascularized fibular graft for bridging large skeletal defects of the upper extremity. *Microsurgery* 31:190–197. doi:10.1002/micr.20862

129. Stevenson S (1998) Enhancement of fracture healing with autogenous and allogeneic bone grafts. *Clin Orthop Relat Res* 355:S239–246
130. Strong DM et al (1996) Immunologic responses in human recipients of osseous and osteochondral allografts. *Clin Orthop Relat Res* 326:107–114
131. Tataru AM, Wong ME, Mikos AG (2014) In vivo bioreactors for mandibular reconstruction. *J Dent Res* 93:1196–1202. doi:[10.1177/0022034514547763](https://doi.org/10.1177/0022034514547763)
132. Tierney CM, Haugh MG, Liedl J, Mulcahy F, Hayes B, O'Brien FJ (2009) The effects of collagen concentration and crosslink density on the biological, structural and mechanical properties of collagen-GAG scaffolds for bone tissue engineering. *J Mech Behav Biomed Mater* 2:202–209. doi:[10.1016/j.jmbbm.2008.08.007](https://doi.org/10.1016/j.jmbbm.2008.08.007)
133. Vail TP, Urbaniak JR (1996) Donor-site morbidity with use of vascularized autogenous fibular grafts. *J Bone Jt Surg Am* 78:204–211
134. Villa MM, Wang L, Huang J, Rowe DW, Wei M (2015) Bone tissue engineering with a collagen-hydroxyapatite scaffold and culture expanded bone marrow stromal cells. *J Biomed Mater Res B Appl Biomater* 103(2):243–253
135. Villemagne T, Bonnard C, Accadbled F, L'Kaissi M, de Billy B, de Gauzy SJ (2011) Intercalary segmental reconstruction of long bones after malignant bone tumor resection using primary methyl methacrylate cement spacer interposition and secondary bone grafting: the induced membrane technique. *J Pediatr Orthop* 31:570–576. doi:[10.1097/BPO.0b013e31821ffa82](https://doi.org/10.1097/BPO.0b013e31821ffa82)
136. Wong TM, Lau TW, Li X, Fang C, Yeung K, Leung F (2014) Masquelet technique for treatment of posttraumatic bone defects. *TheScientificWorldJournal* 2014:710302. doi:[10.1155/2014/710302](https://doi.org/10.1155/2014/710302)
137. Wood MB, Bishop AT (2007) Massive bone defects of the upper limb: reconstruction by vascularized bone transfer. *Hand Clin* 23:49–56. doi:[10.1016/j.hcl.2007.01.002](https://doi.org/10.1016/j.hcl.2007.01.002)
138. Woon CY, Chong KW, Wong MK (2010) Induced membranes—a staged technique of bone-grafting for segmental bone loss: a report of two cases and a literature review. *J Bone Jt Surg Am* 92:196–201. doi:[10.2106/JBJS.I.00273](https://doi.org/10.2106/JBJS.I.00273)
139. Yajima H, Tamai S, Mizumoto S, Ono H (1993) Vascularised fibular grafts for reconstruction of the femur. *J Bone Jt Surg Br* 75:123–128
140. Zappaterra T et al (2011) Induced membrane technique for the reconstruction of bone defects in upper limb. A prospective single center study of nine cases. *Chir Main* 30:255–263. doi:[10.1016/j.main.2011.06.005](https://doi.org/10.1016/j.main.2011.06.005)
141. Zhen P, Hu YY, Luo ZJ, Liu XY, Lu H, Li XS (2010) One-stage treatment and reconstruction of Gustilo Type III open tibial shaft fractures with a vascularized fibular osteoseptocutaneous flap graft. *J Orthop Trauma* 24:745–751. doi:[10.1097/BOT.0b013e3181d88a07](https://doi.org/10.1097/BOT.0b013e3181d88a07)
142. Zigdon-Giladi H, Bick T, Lewinson D, Machtei EE (2015) Co-transplantation of endothelial progenitor cells and mesenchymal stem cells promote neovascularization and bone regeneration. *Clin Implant Dent Relat Res* 17:353–359. doi:[10.1111/cid.12104](https://doi.org/10.1111/cid.12104)